

1. Introduction

Circulating cell-free DNA (cfDNA) contains rich genetic, epigenetic, and structural information for cancer detection, but its highly fragmented nature, low abundance, and high background noise makes reliable detection of circulating tumour DNA (ctDNA) challenging. Conventional double-stranded DNA (dsDNA) workflows rely on end repair and A tailing (ERAT), which lose damaged or ultra-short molecules and introduce artifacts, limiting sensitivity and increasing the limit of detection (LoD).

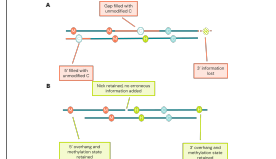


Fig. 1 Molecular impact of double versus single stranded library preparation. (A) Double-stranded ligation and ERAT is error-prone and introduces unmodified cytosines not present in ctDNA. (B) Single-stranded ligation retains original ctDNA features, with no false information introduced through DNA synthesis.

biomodal's duet mosaic workflow is optimised for ctDNA, capturing damaged and ultra-short fragments by ligating adapters directly to single-stranded molecules without ERAT. High material recovery combined with high accuracy 6-base sequencing enables accurate and specific variant and methylation calling, improving reproducibility and lower LoD. duet 6-base mosaic improves the phased multiread readout from the same molecules, including genetic variants, independent 5mC and 5hmC, and native fragmentomic features, reducing false negatives and improving confident detection of ctDNA.



Fig. 2 The duet mosaic workflow provides complete end-to-end solutions for liquid biopsy applications. The workflow includes: the duet mosaic assay, duet software and the modality XPLR software toolkit, designed for accessible, scalable analysis of 5' and 6-base data.

3. duet mosaic captures shorter fragments and maintains high fidelity fragmentomic information

Table 1 Quantification of short ctDNA fragments (below ~145bp)

Short ctDNA fragments are known to be enriched in ctDNA¹. duet 6-base mosaic captures more short fragments than a conventional ERAT workflow.

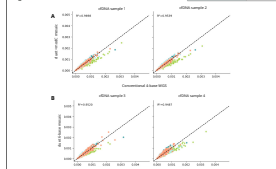


Fig. 5 ctDNA end motif frequencies strongly correlate with WGS 95% agreement between duet +modC mosaic (A) and duet 6-base mosaic (B) and whole-genome sequencing (WGS) from matched clinical ctDNA samples when comparing the frequency of calling A (dark teal), T (coral), G (light teal) or C (green) at the 5' fragment end. With a duet 6-base mosaic you can extend end motifs to be 5mC and 5hmC aware, increasing the biomarkers space from 256 to 4,096, enabling higher granularity and improving classifier performance¹⁹.

2. duet mosaic has market leading methylation and genetic accuracy, including detection of C>T variants

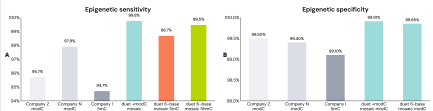
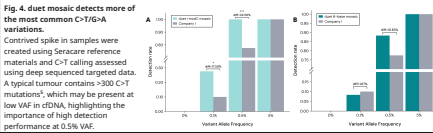


Fig. 3 duet mosaic methylation sensitivity (A) and specificity (B) compared to other epigenetic sequencing technologies. For duet mosaic, sensitivity and specificity are assessed using methylated pUC19 control and unmethylated lambda controls that are included in the every kit. 5hmC sensitivity is calculated using the included short oligo controls. For other sequencing technologies publicly available data was used²⁰. duet mosaic has a 6-9 fold lower number of false positives than Company 1 whilst achieving higher sensitivity to recover the real methylation signal.



4. duet mosaic enables ultra-low LoD through combined genetic and methylation information

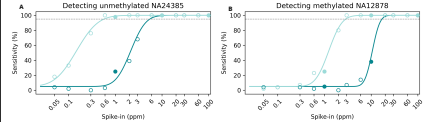


Fig. 6 duet 6-base mosaic LoD is improved through use of integrated genetic and methylation information from the same ctDNA fragment LoD95 was evaluated using Seraseq Sequencing ctDNA Mutation Mix (NA24385) and methylated NA12878 alone or in spike in dilutions (1, 10 and 100 ppm). The two genomes differ at 4,366 homozygous SNVs, targeted using a 1Mb sequencing panel. Experimental dilutions were complemented by 4M-read in-silico mixtures at additional concentrations generated by subsampling. Evidence for methylated NA12878 or unmethylated NA24385 was assessed using joint read-level methylation and SNV information or SNV information alone. LoD95 was estimated by probit regression (95% specificity). Combining genetic and epigenetic signal at single read-level enables an order of magnitude lower LoD compared to genetics alone, enhancing ctDNA detection capabilities.

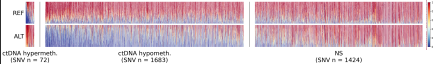


Fig. 7 The majority of SNVs in late stage colorectal cancer (CRC) ctDNA have differential methylation on the same read Tumour-associated SNVs were identified from matched tumour-normal samples from a Stage IV CRC patient (SNV calls and matched ctDNA provided by NeoGenomics). Following 6-base profiling of the ctDNA, reads at each SNV were stratified into ALT (tumour variant indicating a ctDNA fragment) and REF (reference allele, background ctDNA). Read-level methylation differed between ALT and REF molecules at 54% of SNVs, predominantly through hypomethylation on ALT reads (n = 1,683; hypomethylated n = 72). Thus, thousands of loci carry co-occurring genetic and methylation signals on the same ctDNA molecule, providing opportunities for the 6-base genome to improve ctDNA detection and lower LoD versus genetics alone.

5. Conclusions

Sensitive ctDNA detection requires high analytical performance to reliably detect tumour-derived signals at very low prevalence. duet mosaic achieves ultra-low limits of detection by combining high material recovery with outstanding sensitivity and specificity across genomic and epigenetic features:

- Better capture of short ctDNA molecules that are enriched in ctDNA.
- Marker-leading methylation sensitivity, enabling detection of the most challenging biomarkers while maintaining exceptionally low false positive rates.
- Complete SNV detection including common C>T variants.
- The ability to combine common combined genetic and methylation changes on the same DNA fragment to lower LoD by an order of magnitude.

6. References

1. Zhou, X. et al. Epigenetic analysis of cell-free DNA by fragmentomic profiling. *Nat. Adv. Sci.* **2**, 1-10, 2020 (2020).
2. Alizadeh, A. et al. Single-cell whole-genome sequencing of human cells reveals chromatin and genomic architecture in embryonic stem cells. *Nature* **543**, 173-178, 2017.
3. Alizadeh, A. et al. Single-cell whole-genome sequencing of human cells reveals chromatin and genomic architecture in embryonic stem cells. *Nature* **543**, 173-178, 2017.
4. Alizadeh, A. et al. Single-cell whole-genome sequencing of human cells reveals chromatin and genomic architecture in embryonic stem cells. *Nature* **543**, 173-178, 2017.
5. Alizadeh, A. et al. Single-cell whole-genome sequencing of human cells reveals chromatin and genomic architecture in embryonic stem cells. *Nature* **543**, 173-178, 2017.